# Package 'GenomeAdmixR'

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Type Package

Title Simulate Admixture of Genomes

Version 2.1.11

**Description** Individual-based simulations forward in time, simulating how patterns in ancestry along the genome change after admixture. Full description can be found in Janzen (2021) <doi:10.1111/2041-210X.13612>.

License GPL (>= 2)

URL https://github.com/thijsjanzen/GenomeAdmixR

BugReports https://github.com/thijsjanzen/GenomeAdmixR/issues

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Author Thijs Janzen [aut, cre], Fernando Diaz G. [ctb], Richèl J.C. Bilderbeek [ctb]

Maintainer Thijs Janzen <thijsjanzen@gmail.com>

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#### Description

Individual-based simulations forward in time, simulating how patterns in ancestry along the genome change after admixture. The simulation assumes Wright-Fisher dynamics, e.g. random mating and non-overlapping generations. In the simulation, instead of specific alleles, local ancestry is tracked, thus assuming that local molecular data can always be uniquely traced back to one of the founding individuals (populations). The package provides functionality to perform such simulations, but also to perform post-hoc statistical analyses and to visualize the obtained results.

Version 2.1.10 - Fixed memorby bug, improved documentation

Version 2.1.9 - updated tbb::task\_scheduler\_init to tbb::global\_control

Version 2.1.7 - Improve documentation

Version 2.1.6 - check classes with inherits

Version 2.1.5 - Removed debugging output

- Version 2.1.4 Only output when verbose = TRUE
- Version 2.1.3 Changed DOI link in description

Version 2.1.2 - Improved testing

Version 2.1.1 - Removed GNU make dependency

Version 2.1 - Removed error in calculate\_allele\_frequency

Version 2.0.1 - Moved migration outside the modules

Version 2.0 - Added ancestry\_module and sequence\_module to distinguish between implementa-

tions of the model

Version 1.2 - Added example sequencing data

Version 1.2 - Added the option to load sequence data for admixing

Version 1.1 - Fixed a minor bug with plot\_joyplot\_frequencies

Version 1.1 - Improved tests

Version 1.1 - Improved recombination code (again)

Version 1.0 - Release associated with bioRxiv submission, to be found here: https://doi.org/10.1101/2020.10.19.343491

Version 0.66 - Improved recombination code, about twice as fast

Version 0.65 - Added testing and added logo

Version 0.64 - Reduced cyclomatic complexity

Version 0.63 - Updated random number generation

Version 0.62 - Updated to Roxygen

Version 0.61 - Added plot\_over\_time

Version 0.60 - Added admixture with migration

Version 0.59 - Updated frequency code under the hood

Version 0.58 - Renamed to GenomeAdmixR

Version 0.58 - Collapsed and improved many functions

Version 0.57 - Added function to generate admixed individuals

Version 0.56 - Added starting frequencies to 'simulate\_admixture'

Version 0.55 - Extended 'calculate\_marker\_frequency' to handle a vector of locations

Version 0.55 - Increased accuracy of choosing a random position for recombination, this should prevent the rare bug fixed in version 0.54

Version 0.54 - Fixed a MAJOR bug regarding recombination: in rare cases, a crossover position

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could be picked on an existing junction, due to the limited number of digits in uniform()

Version 0.54 - Improved plot\_difference\_frequencies to handle modified input

Version 0.53 - Added multiplicative\_selection

Version 0.52 - Added plot\_difference\_frequencies

Version 0.51 - Added tajima's d calculation

Version 0.50 - Added simulated\_admixture until

Version 0.49 - Added 'simulate' to cpp

Version 0.48 - Added a general 'simulate' function

Version 0.47 - Changed the effect of migration

Version 0.46 - Added joyplot & increase\_ancestor

Version 0.45 - Removed create\_two\_populations

Version 0.44 - Added tracking regions

Version 0.43 - Fixed bugs in select\_population

Version 0.42 - Added initial and final frequency tables

Version 0.41 - Added multiple marker support

Version 0.40 - Collapsed selection functions

Version 0.39 - Added support for non-additive selection

Version 0.38 - Added track frequencies

Version 0.37 - Removed selection on regions

Version 0.36 - Added progress\_bar option

Version 0.35 - Added calculate\_marker\_frequency

Version 0.34 - Added selection\_markers

Version 0.33 - Fixed bugs in selection

Version 0.32 - Moved Fish.h code to Fish.cpp

Version 0.31 - Changed random number generator to R based

Version 0.30 - Added Recombination = 1 code

Version 0.29 - Changed internal junction representation: removed .left

Version 0.28 - Reverted to Agner Fog Random number generation

Version 0.27 - Speed up return types

Version 0.26 - Added class verification code

Version 0.25 - Squashed plotting bug

Version 0.24 - Removed Output.cpp

Version 0.23 - Removed number\_of\_founders from calc\_allele\_spectrum

Version 0.22 - Added save and load functions

Version 0.21 - Changed random-seed management

Version 0.20 - Removed superfluous code

Version 0.19 - Removed number\_of\_founders from Fst and LD code

Version 0.18 - Start of tracking changes

#### Author(s)

Thijs Janzen Maintainer: (thijsjanzen@gmail.com)

#### References

Janzen T, Diaz F. Individual-based simulations of genome evolution with ancestry: The GenomeAdmixR R package. Methods Ecol Evol. 2021; 12: 1346–1357. https://doi.org/10.1111/2041-210X.13612 ancestry\_module

#### Description

Module to perform simulations based on local ancestry

#### Usage

```
ancestry_module(
    input_population = NA,
    number_of_founders = 2,
    initial_frequencies = NA,
    morgan = 1,
    markers = NA,
    track_junctions = FALSE
)
```

#### Arguments

input\_population

Potential earlier simulated population used as starting point for the simulation. If not provided by the user, the simulation starts from scratch.

	······································
number_of_found	lers Number of unique ancestors / ancestries to be tracked in the simulation
initial_frequer	
morgan	vector is normalized. Length of the genomic stretch simulated, expressed in Morgan (e.g. the number of crossovers during meiosis)
markers	A vector of locations of markers, with the location in Morgan. Ancestry at these marker positions is tracked for every generation.
track_junctions	Tracks the average number of junctions over time if TRUE

#### Value

list with type = "Ancestry". Can be used in simulate\_admixture.

calculate\_allele\_frequencies

Calculate allele frequencies

#### Description

Calculate for a number of regularly spaced markers the relative frequency of each ancestor in the population.

#### Usage

```
calculate_allele_frequencies(
  source_pop,
  locations = seq(0, 1, length.out = 100),
  progress_bar = FALSE
)
```

#### Arguments

source_pop	Population for which to estimate allele frequencies
locations	A vector indicating the locations (in Morgan) where to calculate the allele fre- quencies.
progress_bar	Displays a progress_bar if TRUE. Default value is TRUE

#### Details

Markers are equidistantly spaced, with a distance of step\_size in between them.

#### Value

A tibble containing the allele frequencies

#### Examples

calculate\_average\_ld Calculates the ld between two alleles

#### Description

calculate the average ld between two loci

#### Usage

```
calculate_average_ld(alleles_pos_1, alleles_pos_2)
```

#### Arguments

alleles\_pos\_1 alleles at locus 1 alleles\_pos\_2 alleles at locus 2

#### Value

a list with two entries: LD and r\_squared

```
calculate_dist_junctions
```

collect the full distribution of junctions in the population

#### Description

calculates the distribution of junctions across the population

#### Usage

```
calculate_dist_junctions(pop)
```

#### Arguments

pop object of the class 'population'

#### Value

vector with two entries per individual, each indicating the number of junctions in the respective chromosomes

calculate\_fst

#### Description

The FST value between two populations is calculated, given a number of markers. Markers are superimposed upon the (known) ancestry along the chromosome for all sampled individuals. Markers can be chosen to be regularly spaced, or randomly distributed.

#### Usage

```
calculate_fst(
   pop1,
   pop2,
   sampled_individuals = 10,
   number_of_markers = 100,
   random_markers = FALSE
)
```

#### Arguments

pop1	Population object	
pop2	Population object	
sampled_individuals		
	Number of individuals to base the FST upon. Individuals are randomly drawn from each population, a lower number speeds up calculations.	
number_of_markers		
	Number of markers along the chromosome used to calculate FST metrics.	
random_markers	If TRUE, markers are randomly spaced along the chromosome, if FALSE, markers are equidistantly spaced along the chromosome.	

#### Details

Uses the function wc from the package hierfstat to calculate the FST. The function wc computes the Weir and Cockerham F statistic.

#### Value

FST value

#### Examples

two\_populations <- simulate\_admixture(</pre>

```
module = ancestry_module(),
migration = migration_settings(migration_rate = 0.01,
population_size = c(100, 100)))
```

calculate\_heterozygosity

Calculate heterozygosity

#### Description

Calculate the average population level heterozygosity

#### Usage

```
calculate_heterozygosity(source_pop, locations, progress_bar = FALSE)
```

#### Arguments

source_pop	Population for which to estimate allele frequencies, or a list of individuals for which to calculate average heterozygosity
locations	A vector indicating the locations (in Morgan) of markers for which to calculate the heterozygosity
progress_bar	Displays a progress_bar if TRUE. Default value is TRUE

#### Value

A tibble containing the heterozygosities

calculate_ld	Calculate linkage disequilibrium statistics This function calculates
	two matrices, once containing all pairwise linkage disequilibrium (ld)
	values, and one matrix containing all pairwise r statistics

#### Description

Calculate linkage disequilibrium statistics This function calculates two matrices, once containing all pairwise linkage disequilibrium (ld) values, and one matrix containing all pairwise r statistics

#### Usage

```
calculate_ld(pop, sampled_individuals = 10, markers = NA, verbose = FALSE)
```

#### Arguments

рор	focal population
sampled_individuals	
	Number of individuals randomly sampled to calculate the LD matrices
markers	vector of markers. If a single number is used, that number of markers is ran- domly placed along the genome.
verbose	display verbose output, default is FALSE.

#### Value

An object containing two items:

ld_matrix	Pairwise ld statistics for all markers
rsq_matrix	Pairwise rsq statistics for all markers

#### Examples

```
ylab = "Linkage Disequilibrium")
```

calculate\_marker\_frequency

Calculate allele frequencies at a specific marker location

#### Description

Calculate the relative frequency of each ancestor in the population at a specific marker location

#### Usage

```
calculate_marker_frequency(pop, location)
```

#### Arguments

рор	Population for which to estimate allele frequencies at the given marker
location	A vector or scalar of location(s) along the chromosome for which allele frequen-
	cies are to be calculated. Locations are in Morgan.

#### Value

A tibble containing the frequency of each present ancestor at the provided location. Ancestors with frequency = 0 are dropped out of the table. The tibble contains three columns: location, ancestor and frequency.

#### Examples

```
combine_input_data combine sequence data that was previously read from file into a popu-
lation
```

#### Description

Create data in a format that can be used by GenomeAdmixR, entries are sampled randomly from each input data set, with replacement. Probability of sampling from each input data set is driven by the input frequencies, and total number of individuals sampled is driven by pop\_size.

#### Usage

```
combine_input_data(input_data_list, frequencies = NA, pop_size)
```

#### Arguments

input_data_list		
	list where each entry is the result of create_input_data	
frequencies	frequency of each entry in the list in the starting population	
pop_size	intended population size	

#### Value

the input data entries are combined to one single population that can be used to seed simulate\_admixture\_data. Output is identical to create\_input\_data

```
create_artificial_genomeadmixr_data
```

function to generate artificial genomeadmixr\_data

#### Description

function to generate artificial genomeadmixr\_data

#### Usage

```
create_artificial_genomeadmixr_data(
   number_of_individuals,
   marker_locations,
   used_nucleotides = 1:4,
   nucleotide_frequencies = NA
)
```

#### Arguments

number\_of\_individuals number of individuals marker\_locations location of markers, either in bp or Morgan used\_nucleotides subset or full set of [1/2/3/4] (reflecting a/c/t/g) nucleotide\_frequencies frequencies of the used nucleotides, if not provided, equal frequencies are assumed.

#### Value

genomeadmixr\_data object ready for simulate\_admixture\_data

create\_iso\_female *function to simulate creation of an isofemale line* 

#### Description

create\_isofemale simulates the creation of an isofemale line through extreme inbreeding.

#### dgrp2.3R.5k.data

#### Usage

```
create_iso_female(
  module = ancestry_module(),
  n = 1,
  inbreeding_pop_size = 100,
  run_time = 2000,
  num_threads = 1,
  verbose = FALSE
)
```

#### Arguments

module	Source population from which isofemales are generated
n	Number of isofemales to be generated
inbreeding_pop_size	
	Population size of the population used to generate homozygous individuals
run_time	Maximum runtime used for inbreeding
num_threads	number of threads. Default is 1. Set to -1 to use all available threads
verbose	Displays verbose output if TRUE. Default value is FALSE

#### Details

To create an isofemale, two individuals are randomly picked from the source population. Using these two individuals, a new population is seeded, of size inbreeding\_pop\_size. Then, this population is allowed to inbreed until either run\_time is reached, or until all individuals are homozygous and genetically identical, whatever happens first.

#### Value

A list of length n, where each entry is a fully homozygous isofemale.

dgrp2.3R.5k.data A subset of sequencing data from the Drosophila Genetics Reference Panel

#### Description

This data set contains sequences from the 3R chromosome. Included are 4603 SNPs with at least 0.05 minor allele frequency, sequenced across 410 isofemale lines. Sequences were downloaded from <a href="http://dgrp2.gnets.ncsu.edu/data.html">http://dgrp2.gnets.ncsu.edu/data.html</a>.

#### Usage

data("dgrp2.3R.5k.data")

#### Format

genomeadmixr\_data object

#### References

Mackay, T., Richards, S., Stone, E. et al. The Drosophila melanogaster Genetic Reference Panel. Nature 482, 173–178 (2012). <a href="https://doi.org/10.1038/nature10811">https://doi.org/10.1038/nature10811</a>

#### Examples

```
data("dgrp2.3R.5k.data")
simulate_admixture(
    module = sequence_module(molecular_data = dgrp2.3R.5k.data),
    pop_size = 100,
    total_runtime = 10)
```

iso\_female\_ancestry Create isofemale

#### Description

Creates isofemale individuals, given a population

#### Usage

```
iso_female_ancestry(
  source_pop = NA,
  n = 1,
  inbreeding_pop_size = 100,
  run_time = 2000,
  morgan = 1,
  num_threads = 1,
  verbose = FALSE
)
```

#### Arguments

source_pop	Source population from which isofemales are generated
n	Number of isofemales to be generated
inbreeding_pop	_size
	Population size of the population used to generate homozygous individuals
run_time	Maximum runtime used for inbreeding
morgan	Size of the chromosome in Morgan (e.g. the number of crossovers during meiosis)
num_threads	number of threads. Default is 1. Set to -1 to use all available threads
verbose	Displays verbose output if TRUE. Default value is FALSE

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#### Details

To create an isofemale, two individuals are randomly picked from the source population. Using these two individuals, a new population is seeded, of size inbreeding\_pop\_size. Then, this population is allowed to inbreed until either run\_time is reached, or until all individuals are homozygous and genetically identical, whatever happens first.

#### Value

A list of length n, where each entry is a fully homozygous isofemale.

iso\_female\_sequence Create isofemale

#### Description

Creates isofemale individuals, given a population

#### Usage

```
iso_female_sequence(
    input_data = NA,
    n = 1,
    inbreeding_pop_size = 100,
    run_time = 2000,
    morgan = 1,
    recombination_rate = NA,
    num_threads = 1,
    verbose = FALSE
)
```

#### Arguments

input_data	Source population from which isofemales are generated	
n	Number of isofemales to be generated	
inbreeding_pop	_size	
	Population size of the population used to generate homozygous individuals	
run_time	Maximum runtime used for inbreeding	
morgan	Size of the chromosome in Morgan (e.g. the number of crossovers during meio-	
	sis)	
recombination_rate		
	rate in cM / Mbp, used to map recombination to the markers. If the recombination_rate is not set, the value for Morgan is used, assuming that the markers included span an entire chromosome.	
num_threads	number of threads. Default is 1. Set to -1 to use all available threads	
verbose	Displays verbose output if TRUE. Default value is FALSE	

#### Details

To create an isofemale, two individuals are randomly picked from the source population. Using these two individuals, a new population is seeded, of size inbreeding\_pop\_size. Then, this population is allowed to inbreed until either run\_time is reached, or until all individuals are homozygous and genetically identical, whatever happens first.

#### Value

A list of length n, where each entry is a fully homozygous isofemale.

load\_population Load a population from file

#### Description

Loads a population that has previously been written to file.

#### Usage

```
load_population(file_name)
```

#### Arguments

file\_name Name of the file to save the population

#### Details

This function is a wrapper for readRDS.

#### Value

A population object

#### See Also

save\_population

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migration\_settings Function to manage settings associated with migration

#### Description

creates a list with settings associated with migration.

#### Usage

```
migration_settings(
  migration_rate = NA,
  stop_at_critical_fst = FALSE,
  critical_fst = NA,
  population_size = c(100, 100),
  initial_frequencies = list(c(1, 0), c(0, 1)),
  generations_between_update = 10,
  sampled_individuals = 10,
  number_of_markers = 100,
  random_markers = TRUE
)
```

#### Arguments

migration_rate	Rate of migration between the two populations. Migration is implemented such that with probability m (migration rate) one of the two parents of a new offspring
	is from the other population, with probability 1-m both parents are of the focal population.
stop_at_critica	l_fst
	option to stop at a critical FST value, default is FALSE
critical_fst	the critical fst value to stop, if stop_simulation_at_critical_fst is TRUE
population_size	
	vector of population sizes, one size for each population
initial_frequen	cies
	A list describing the initial frequency of each ancestor in each population. Each entry in the list contains a vector with the frequencies for all ancestor. The length
	of the vector indicates the number of unique ancestors. If a vector not summing to 1 is provided, the vector is normalized.
generations_bet	•
80	The number of generations after which the simulation has to check again whether the critical Fst value is exceeded
sampled_individ	
	Number of individuals to be sampled at random from the population to estimate
	Fst
number_of_marke	rs
	Number of markers to be used to estimate Fst
random_markers	Are the markers to estimate Fst randomly distributed, or regularly distributed? Default is TRUE.

#### Value

list with migration associated settings. To be used to pass on migration settings to simulate\_admixture.

```
plink_to_genomeadmixr_data
```

function to convert plink style (ped/map) data to genome\_admixr\_data

#### Description

function to convert plink style (ped/map) data to genome\_admixr\_data

#### Usage

```
plink_to_genomeadmixr_data(
   ped_data,
   map_data,
   chosen_chromosome,
   verbose = FALSE
)
```

#### Arguments

ped_data	result of read.table(ped_file, header = F)	
map_data	result of read.table(map_file, header = F)	
chosen_chromosome		
	chromosome of choice	
verbose	verbose output	

#### Value

genomeadmixr\_data object ready for simulate\_admixture\_data

plot.individual plot the genome of an individual

### Description

visualise ancestry blocks on both chromosomes

#### Usage

```
## S3 method for class 'individual'
plot(x, cols = NA, ...)
```

#### plot\_chromosome

#### Arguments

х	object of type individual
cols	colors for the different ancestors
	other arguments

#### Value

No return value

plot_chromosome	e plots a chromosome	

#### Description

This function plots a chromosome in the range [xmin, xmax]. Colors indicate different ancestry.

#### Usage

plot\_chromosome(chrom, xmin = 0, xmax = 1)

#### Arguments

chrom	object of type chromosome, typically a table with two columns. The first column indicates the start of an ancestry block (location in Morgan), the second column indicates the ancestry type.
xmin	minimum value of the range, $default = 0$ .
xmax	maximum value of the range, $default = 1$ .

#### Value

No return value

#### Examples

```
wildpop = simulate_admixture(
    module = ancestry_module(number_of_founders = 10, morgan = 1),
    pop_size = 1000,
    total_runtime = 10)
```

plot\_chromosome(chrom = isofemale[[1]]\$chromosome1)

```
# and a detail of the chromosome:
plot_chromosome(chrom = isofemale[[1]]$chromosome1,
                xmin = 0.4,
                xmax = 0.6)
```

```
plot_difference_frequencies
```

Plot the change in frequency between the start and end of a simulation

#### Description

This function plots the change in frequency of one or multiple ancestors after performing a simulation.

#### Usage

```
plot_difference_frequencies(
  results,
  picked_ancestor = "ALL",
  picked_population = 1
)
```

#### Arguments

results

An object which is the result of simulate\_admixture being a list with four properties: population, frequencies, initial\_frequencies and final frequencies

```
picked_ancestor
```

Default is "ALL", where different colors indicate different ancestors. Alternatively, for clarity, the user can specify a specific ancestral allele, and only that allele is plotted

```
picked_population
```

If multiple populations were simulated (in the case of simulate\_admixture\_migration), which population should be plotted? Default is population\_1.

#### Value

a ggplot2 object

#### Examples

```
s <- 0.1
select_matrix <- matrix(nrow = 1, ncol = 5)</pre>
select_matrix[1, ] <- c(0.25, 1.0, 1 + 0.5 * s, 1 + s, 0)</pre>
markers <- seq(from = 0.2, to = 0.3, length.out = 100)
selected_pop <- simulate_admixture(</pre>
                     module = ancestry_module(number_of_founders = 10,
```

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```
morgan = 1,
markers = markers),
pop_size = 1000,
total_runtime = 11,
select_matrix = select_matrix)
require(ggplot2)
plot_difference_frequencies(results = selected_pop,
picked_ancestor = "ALL")
```

plot\_dist\_junctions plot the distribution of junctions

#### Description

plots the distribution of junctions in the population using base R

#### Usage

```
plot_dist_junctions(pop)
```

#### Arguments

pop of the class 'population'

#### Value

No return value

plot\_frequencies *Plot the frequencies of all ancestors along the genome.* 

#### Description

This function plots the frequency of all ancestors after performing a simulation.

#### Usage

```
plot_frequencies(
   result,
   locations = seq(0, 1, length.out = 100),
   progress_bar = FALSE
)
```

#### Arguments

result	An object which is the result of select_population or create_population_selection, being a list with four properties: population, frequencies, initial_frequencies and final frequencies
locations	A vector indicating the locations (in Morgan) where to calculate the allele fre- quencies.
progress_bar	Displays a progress_bar if TRUE. Default value is FALSE

#### Value

a ggplot2 object

#### Examples

```
pop <- simulate_admixture(</pre>
             module = ancestry_module(number_of_founders = 4),
             pop_size = 1000,
             total_runtime = 11)
require(ggplot2)
plot_frequencies(result = pop)
```

```
plot_joyplot_frequencies
```

make a joy plot of the distribution of allele frequencies within a region

#### Description

This function plots the distribution of allele frequencies within a region over time, making use of a 'joyplot'

#### Usage

```
plot_joyplot_frequencies(
  frequencies,
  time_points,
 picked_ancestor = "ALL",
 picked_population = 1
)
```

frequencies	A tibble containing four columns: time, location, ancestor, frequency.	
Typically one of the items returned by create_population_selection or select_		
	when the user specifies track_frequency.	
time_points	A sequence of time points for which the user wants to create the joyplot	

Arguments

```
picked_ancestor
```

Default is "ALL", where different colors indicate different ancestors. Alternatively, for clarity, the user can specify a specific ancestral allele, and only that allele is plotted

picked\_population

If multiple populations were simulated (in the case of simulate\_admixture\_migration), which population should be plotted? Default is population\_1.

#### Value

a ggplot object

#### Examples

```
s <- 0.01
select_matrix <- matrix(nrow = 1, ncol = 5)</pre>
select_matrix[1, ] <- c(0.25, 1.0, 1 + 0.5 * s, 1 + s, 0)</pre>
markers <- seq(from = 0.2, to = 0.3, length.out = 100)
selected_pop <- simulate_admixture(</pre>
                    module = ancestry_module(number_of_founders = 10,
                                              morgan = 1,
                                              markers = markers),
                    pop_size = 1000,
                    total_runtime = 11,
                    select_matrix = select_matrix)
require(ggplot2)
plot_joyplot_frequencies(frequencies = selected_pop$frequencies,
                          time_points = 0:11,
                          picked_ancestor = "ALL")
# joyplot frequencies returns a ggplot object, so we can
# add extra elements:
plot_joyplot_frequencies(frequencies = selected_pop$frequencies,
                          time_points = 0:11,
                         picked_ancestor = "ALL") +
 ggplot2::xlab("Location") +
 ggplot2::ylab("Generations")
```

plot\_over\_time Plot the frequencies of all ancestors over time

#### Description

This function plots the frequency of all ancestors over time at a specific location on the chromosome, after performing a simulation.

#### Usage

```
plot_over_time(frequencies, focal_location)
```

#### Arguments

frequencies A tibble containing four columns: time, location, ancestor, frequency. A fifth colum population can be included if the tibble is the result of simulate\_admixture\_migration.

focal\_location Location (in Morgan) where to plot the allele frequencies.

#### Value

a ggplot2 object

#### Examples

```
pop <- simulate_admixture(
                module = ancestry_module(number_of_founders = 10,
                     markers = 0.5),
                pop_size = 1000,
                total_runtime = 11)
require(ggplot2)
plot_over_time(frequencies = pop$frequencies,
                    focal_location = 0.5)</pre>
```

plot\_start\_end Plot both the starting frequencies and the final frequencies in one plot

#### Description

This function plots the distribution of both the starting and the final frequencies in one plot

#### Usage

```
plot_start_end(results, picked_ancestor = "ALL", picked_population = 1)
```

#### Arguments

results	An object which is the result of simulate_admixture, being a list with four properties: population, frequencies, initial_frequencies and final frequencies	
picked_ancestor		
	Default is "ALL", where different colors indicate different ancestors. Alterna- tively, for clarity, the user can specify a specific ancestral allele, and only that allele is plotted	
picked_population		
	If multiple populations were simulated (in the case of simulate_admixture_migration), which population should be plotted? Default is population_1.	

#### Value

a ggplot object

#### Examples

print.genomeadmixr\_data

```
print an individual to the console
```

#### Description

prints an object of class genomeadmixr\_data to the console

#### Usage

```
## S3 method for class 'genomeadmixr_data'
print(x, ...)
```

#### Arguments

х	individual
	other arguments

#### Value

No return value

print.individual print an individual to the console

#### Description

prints an object of class individual to the console

#### Usage

```
## S3 method for class 'individual'
print(x, ...)
```

#### Arguments

х	individual
	other arguments

#### Value

No return value

print.population print a population object

#### Description

prints the contents of a population nicely

#### Usage

```
## S3 method for class 'population'
print(x, ...)
```

#### Arguments

х	input population
•••	other arguments

### Value

No return value

read\_input\_data read sequence data from file to be used in simulation

#### Description

Create data in a format that can be used by GenomeAdmixR

#### Usage

```
read_input_data(
   file_names,
   type,
   chosen_chromosome,
   number_of_snps = NA,
   random_snps = TRUE,
   verbose = FALSE
)
```

#### Arguments

file_names	names of input files	
type	type of data, options are 'ped' and 'vcf'	
chosen_chromosome		
	GenomeAdmixR simulates only a single chromosome.	
number_of_snps	number of snps to be loaded from file, default is to load all snps	
random_snps	if a subset of all snps has to be taken, should these be sampled sequentially (e.g. the first 100 snps) or randomly (100 randomly sampled snps) (examples are for 'number_of_snps' = 100).	
verbose	give verbose output	

#### Value

list with two properties: genomes a matrix with the sequence translated to numerics, such that [actg] corresponds to [1234], and missing data is represented with "-". Rows in the matrix correspond to chromosomes, and columns represent bases. Two consecutive rows represent an individual, such that rows 1-2 are individual, rows 3-4 are one individual etc. markers corresponds to the locations of the markers (in bp) on the chosen chromosome.

save\_population Save a

#### Description

Saves a population to file for later use

#### Usage

save\_population(population, file\_name, compression = TRUE)

#### Arguments

population	Object of class population
file_name	Name of the file to save the population
compression	By default, the population is compressed to reduce file size. See for more infor- mation saveRDS

#### Details

This function functions as a wrapper for the base function saveRDS.

#### Value

No return value

sequence\_module create sequence module

#### Description

creates a sequence module, which contains all relevant information in order to perform a simulation based on sequence data.

#### Usage

```
sequence_module(
  molecular_data = NA,
  initial_frequencies = NA,
  morgan = 1,
  recombination_rate = NA,
  markers = NA,
  mutation_rate = 0,
  substitution_matrix = matrix(1/4, 4, 4)
)
```

#### Arguments

molecular_data	Genomic data used as input, should be of type genomeadmixr_data. Either a single dataset is provided, or a list of multiple genomeadmixr_data objects.	
initial_frequencies		
	A vector describing the initial contribution of each provided input data set to the starting hybrid swarm. By default, equal frequencies are assumed. If a vector not summing to 1 is provided, the vector is normalized.	
morgan	Length of the molecular sequence in Morgan (e.g. the number of crossovers during meiosis), alternatively, the recombination rate can be used, see below.	
recombination_rate		
	rate in $cM / Mbp$ , used to map recombination to the markers. If the recombination_rate is not set, the value for Morgan is used, assuming that the markers included span an entire chromosome.	
markers	A vector of locations of markers, these markers are tracked for every generation.	
mutation_rate	the per base probability of mutation. Default is 0.	
substitution_matrix		
	a 4x4 matrix representing the probability of mutating to another base (where $[1/2/3/4] = [a/c/t/g]$ ), conditional on the event of a mutation happening. Default is the JC69 matrix, with equal probabilities for all transitions / transversions.	

#### Value

sequence module object, used as starting point for simulate\_admixture.

simulate_admixture	Individual based simulation of the breakdown of contiguous ancestry blocks.

#### Description

Individual based simulation of the breakdown of contiguous ancestry blocks, with or without selection. Simulations can be started from scratch, or from a predefined input population.

#### Usage

```
simulate_admixture(
  module = ancestry_module(),
  pop_size = 100,
  total_runtime = 100,
  migration = migration_settings(),
  select_matrix = NA,
  multiplicative_selection = TRUE,
  verbose = FALSE,
  num_threads = 1
)
```

#### Arguments

module	$Chosen \ module \ to \ simulate, \ either \ created \ with \ module\_ancestry \ or \ module\_sequence.$	
pop_size	The number of individuals in the population. If the number is larger than the number of individuals in the input population (if provided), additional individuals are sampled randomly from the input population to reach the intended size.	
total_runtime	Number of generations	
migration	settings associated with migration, should be created with migration_settings	
select_matrix	Selection matrix indicating the markers which are under selection. If not pro- vided by the user, the simulation proceeds neutrally. If provided, each row in the matrix should contain five entries:	
	• location of the marker under selection (in Morgan)	
	• fitness of wildtype (aa)	
	• fitness of heterozygote (aA)	
	• fitness of homozygote mutant (AA)	
	• Ancestral type that represents the mutant allele A	
multiplicative_selection		
	Default: TRUE. If TRUE, fitness is calculated for multiple markers by multi- plying fitness values for each marker. If FALSE, fitness is calculated by adding fitness values for each marker.	
verbose	Verbose output if TRUE. Default value is FALSE	
num_threads	number of threads. Default is 1. Set to -1 to use all available threads	

#### Value

A list with: population a population object, and three tibbles with allele frequencies (only contain values of a vector was provided to the argument markers: frequencies, initial\_frequencies and final\_frequencies. Each tibble contains four columns, time, location, ancestor and frequency, which indicates the number of generations, the location along the chromosome of the marker, the ancestral allele at that location in that generation, and finally, the frequency of that allele.

#### Examples

```
recombination_rate = 0.2,
    mutation_rate = 1e-5),
pop_size = 1000,
total_runtime = 10)
```

simulate\_ancestry Individual based simulation of the breakdown of contiguous ancestry blocks.

#### Description

Individual based simulation of the breakdown of contiguous ancestry blocks, with or without selection. Simulations can be started from scratch, or from a predefined input population.

#### Usage

```
simulate_ancestry(
    input_population = NA,
    pop_size = NA,
    number_of_founders = 2,
    initial_frequencies = NA,
    total_runtime = 100,
    morgan = 1,
    num_threads = 1,
    select_matrix = NA,
    markers = NA,
    verbose = FALSE,
    track_junctions = FALSE,
    multiplicative_selection = TRUE
)
```

#### Arguments

input_population		
	Potential earlier simulated population used as starting point for the simulation. If not provided by the user, the simulation starts from scratch.	
pop_size	The number of individuals in the population. If the number is larger than the number of individuals in the input population (if provided), additional individuals are sampled randomly from the input population to reach the intended size.	
number_of_founders		
	Number of unique ancestors	
initial_frequencies		
	A vector describing the initial frequency of each ancestor. By default, equal frequencies are assumed. If a vector not summing to 1 is provided, the vector is normalized.	
total runtime	Number of generations	

total\_runtime Number of generations

morgan	Length of the chromosome in Morgan (e.g. the number of crossovers during meiosis)
num_threads	number of threads. Default is 1. Set to -1 to use all available threads
select_matrix	Selection matrix indicating the markers which are under selection. If not pro- vided by the user, the simulation proceeds neutrally. If provided, each row in the matrix should contain five entries:
	• location of the marker under selection (in Morgan)
	• fitness of wildtype (aa)
	• fitness of heterozygote (aA)
	• fitness of homozygote mutant (AA)
	• Ancestral type that represents the mutant allele A
markers	A vector of locations of markers (relative locations in [0, 1]). If a vector is provided, ancestry at these marker positions is tracked for every generation.
verbose	Verbose output if TRUE. Default value is FALSE
track_junctions	
	Track the average number of junctions over time if TRUE
multiplicative_selection	
	Default: TRUE. If TRUE, fitness is calculated for multiple markers by multiplying fitness values for each marker. If FALSE, fitness is calculated by adding fitness values for each marker.

#### Value

A list with: population a population object, and three tibbles with allele frequencies (only contain values of a vector was provided to the argument markers: frequencies, initial\_frequencies and final\_frequencies. Each tibble contains four columns, time, location, ancestor and frequency, which indicates the number of generations, the location along the chromosome of the marker, the ancestral allele at that location in that generation, and finally, the frequency of that allele.

simulate\_ancestry\_migration

Individual based simulation of the breakdown of contiguous ancestry blocks in two populations linked by migration

#### Description

Individual based simulation of the breakdown of contiguous ancestry blocks, with or without selection. Simulations can be started from scratch, or from a predefined input population. Two populations are simulated, connected by migration

#### Usage

```
simulate_ancestry_migration(
 input_population_1 = NA,
  input_population_2 = NA,
 pop_size = c(100, 100),
 initial_frequencies = list(c(1, 0), c(0, 1)),
  total_runtime = 100,
 morgan = 1,
 num_threads = 1,
 select_matrix = NA,
 markers = NA,
 verbose = FALSE,
 track_junctions = FALSE,
 multiplicative_selection = TRUE,
 migration_rate = 0,
 stop_at_critical_fst = FALSE,
 critical_fst = 0.1,
 generations_between_update = 100,
 sampled_individuals = 10,
 number_of_markers = 100,
 random_markers = TRUE
)
```

#### Arguments

input_population_1		
	Potential earlier simulated population used as starting point for the simulation. If not provided by the user, the simulation starts from scratch.	
input_population	on_2	
	Potential earlier simulated population used as starting point for the simulation. If not provided by the user, the simulation starts from scratch.	
pop_size	Vector containing the number of individuals in both populations.	
initial_frequer	ncies	
	A list describing the initial frequency of each ancestor in each population. Each entry in the list contains a vector with the frequencies for all ancestor. The length of the vector indicates the number of unique ancestors. If a vector not summing to 1 is provided, the vector is normalized.	
total_runtime	Number of generations	
morgan	Length of the chromosome in Morgan (e.g. the number of crossovers during meiosis)	
num_threads	number of threads. Default is 1. Set to -1 to use all available threads	
select_matrix	Selection matrix indicating the markers which are under selection. If not provided by the user, the simulation proceeds neutrally. If provided, each row in the matrix should contain five entries:	
	• location of the marker under selection (in Morgan)	

• fitness of wildtype (aa)

	• fitness of hotomorphics $(a A)$	
	• fitness of heterozygote (aA)	
	• fitness of homozygote mutant (AA)	
	• Ancestral type that represents the mutant allele A	
markers	A vector of locations of markers (relative locations in [0, 1]). If a vector is provided, ancestry at these marker positions is tracked for every generation.	
verbose	Verbose output if TRUE. Default value is FALSE	
track_junctions	5	
	Track the average number of junctions over time if TRUE	
multiplicative_	_selection	
	Default: TRUE. If TRUE, fitness is calculated for multiple markers by multi- plying fitness values for each marker. If FALSE, fitness is calculated by adding fitness values for each marker.	
migration_rate	Rate of migration between the two populations. Migration is implemented such that with probability m (migration rate) one of the two parents of a new offspring is from the other population, with probability 1-m both parents are of the focal population.	
<pre>stop_at_critica</pre>	al_fst	
	option to stop at a critical FST value, default is FALSE	
critical_fst	the critical fst value to stop, if stop_simulation_at_critical_fst is TRUE	
generations_bet	tween_update	
	The number of generations after which the simulation has to check again whether the critical Fst value is exceeded	
sampled_individuals		
	Number of individuals to be sampled at random from the population to estimate Fst	
number_of_markers		
	Number of markers to be used to estimate Fst	
random_markers	Are the markers to estimate Fst randomly distributed, or regularly distributed? Default is TRUE.	

#### Value

A list with: population\_1, population\_2 two population objects, and three tibbles with allele frequencies (only contain values of a vector was provided to the argument markers: frequencies, initial\_frequencies and final\_frequencies. Each tibble contains five columns, time, location, ancestor, frequency and population, which indicates the number of generations, the location along the chromosome of the marker, the ancestral allele at that location in that generation, the frequency of that allele and the population in which it was recorded (1 or 2). If a critical fst value was used to terminate the simulation, and object FST with the final FST estimate is returned as well.

 $simulate\_sequence$ 

#### Description

Individual based simulation of the breakdown of contiguous ancestry blocks, with or without selection. Simulations can be started from scratch, or from a predefined input population.

#### Usage

```
simulate_sequence(
    input_data = NA,
    pop_size = NA,
    initial_frequencies = NA,
    total_runtime = 100,
    morgan = 1,
    recombination_rate = NA,
    num_threads = 1,
    select_matrix = NA,
    markers = NA,
    verbose = FALSE,
    multiplicative_selection = TRUE,
    mutation_rate = 0,
    substitution_matrix = matrix(1/4, 4, 4)
)
```

#### Arguments

input_data	Genomic data used as input, should be of type genomeadmixr_data. Either a single dataset is provided, or a list of multiple genomeadmixr_data objects.	
pop_size	Vector containing the number of individuals in both populations.	
initial_frequen	cies	
	A vector describing the initial contribution of each provided input data set to the starting hybrid swarm. By default, equal frequencies are assumed. If a vector not summing to 1 is provided, the vector is normalized.	
total_runtime	Number of generations	
morgan	Length of the chromosome in Morgan (e.g. the number of crossovers during meiosis)	
recombination_rate		
	rate in cM / Mbp, used to map recombination to the markers. If the recombination_rate is not set, the value for Morgan is used, assuming that the markers included span an entire chromosome.	
num_threads	number of threads. Default is 1. Set to -1 to use all available threads	

select_matrix	Selection matrix indicating the markers which are under selection. If not provided by the user, the simulation proceeds neutrally. If provided, each row in the matrix should contain five entries:	
	• location of the marker under selection (in Morgan)	
	• fitness of wildtype (aa)	
	• fitness of heterozygote (aA)	
	• fitness of homozygote mutant (AA)	
	Ancestral type that represents the mutant allele A	
markers	A vector of locations of markers (relative locations in $[0, 1]$ ). If a vector is provided, ancestry at these marker positions is tracked for every generation.	
verbose	Verbose output if TRUE. Default value is FALSE	
multiplicative_selection		
	Default: TRUE. If TRUE, fitness is calculated for multiple markers by multiplying fitness values for each marker. If FALSE, fitness is calculated by adding fitness values for each marker.	
mutation_rate	the per base probability of mutation. Default is 0.	
substitution_matrix		
	a 4x4 matrix representing the probability of mutating to another base (where $[1/2/3/4] = [a/c/t/g]$ ), conditional on the event of a mutation happening. Default is the JC69 matrix, with equal probabilities for all transitions / transversions.	

#### Value

A list with: population a population object, and three tibbles with allele frequencies (only contain values of a vector was provided to the argument markers: frequencies, initial\_frequencies and final\_frequencies. Each tibble contains four columns, time, location, ancestor and frequency, which indicates the number of generations, the location along the chromosome of the marker, the ancestral allele at that location in that generation, and finally, the frequency of that allele.

simulate\_sequence\_migration

Individual based simulation of the breakdown of contiguous ancestry blocks in two populations linked by migration

#### Description

Individual based simulation of the breakdown of contiguous ancestry blocks, with or without selection. Simulations can be started from scratch, or from a predefined input population. Two populations are simulated, connected by migration

#### Usage

```
simulate_sequence_migration(
 input_data_population_1 = NA,
  input_data_population_2 = NA,
 pop_size = c(100, 100),
 total_runtime = 100,
 morgan = 1,
 recombination_rate = NA,
 num_threads = 1,
 select_matrix = NA,
 markers = NA,
 verbose = FALSE,
 multiplicative_selection = TRUE,
 migration_rate = 0,
 stop_at_critical_fst = FALSE,
 critical_fst = NA,
 generations_between_update = 100,
 sampled_individuals = 10,
 number_of_markers = 100,
 random_markers = TRUE,
 mutation_rate = 0,
 substitution_matrix = matrix(1/4, 4, 4)
)
```

#### Arguments

input_data_population_1		
Genomic data used as input, should be created by the function create_input_data or by the function combine_input_data		
ulation_2		
Genomic data used as input, should be created by the function <code>create_input_data</code> or by the function <code>combine_input_data</code>		
Vector containing the number of individuals in both populations.		
Number of generations		
Length of the chromosome in Morgan (e.g. the number of crossovers during meiosis)		
recombination_rate		
rate in cM / Mbp, used to map recombination to the markers. If the recombination_rate is not set, the value for morgan is used, assuming that the markers included span an entire chromosome.		
number of threads. Default is 1. Set to -1 to use all available threads		
Selection matrix indicating the markers which are under selection. If not pro- vided by the user, the simulation proceeds neutrally. If provided, each row in the matrix should contain five entries:		
<ul><li>location of the marker under selection (in Morgan)</li><li>fitness of wildtype (aa)</li></ul>		

	• fitness of heterozygote (aA)		
	• fitness of homozygote mutant (AA)		
	• Ancestral type that represents the mutant allele A		
markers	A vector of locations of markers (relative locations in [0, 1]). If a vector is provided, ancestry at these marker positions is tracked for every generation.		
verbose	Verbose output if TRUE. Default value is FALSE		
multiplicative_selection			
	Default: TRUE. If TRUE, fitness is calculated for multiple markers by multi- plying fitness values for each marker. If FALSE, fitness is calculated by adding fitness values for each marker.		
migration_rate	Rate of migration between the two populations. Migration is implemented such that with probability m (migration rate) one of the two parents of a new offspring is from the other population, with probability 1-m both parents are of the focal population.		
<pre>stop_at_critica</pre>	<pre>stop_at_critical_fst</pre>		
	option to stop at a critical FST value , default is FALSE		
critical_fst	the critical fst value to stop, if stop_simulation_at_critical_fst is TRUE		
generations_bet	generations_between_update		
	The number of generations after which the simulation has to check again whether the critical Fst value is exceeded		
sampled_individ	sampled_individuals		
	Number of individuals to be sampled at random from the population to estimate Fst		
number_of_marke	ers		
	Number of markers to be used to estimate Fst		
random_markers	Are the markers to estimate Fst randomly distributed, or regularly distributed? Default is TRUE.		
mutation_rate	the per base probability of mutation. Default is 0.		
substitution_ma	atrix		
	a 4x4 matrix representing the probability of mutating to another base (where $[1/2/3/4] = [a/c/t/g]$ ), conditional on the event of a mutation happening. Default is the JC69 matrix, with equal probabilities for all transitions / transversions.		

#### Value

A list with: population\_1, population\_2 two population objects, and three tibbles with allele frequencies (only contain values of a vector was provided to the argument markers: frequencies, initial\_frequencies and final\_frequencies. Each tibble contains five columns, time, location, ancestor, frequency and population, which indicates the number of generations, the location along the chromosome of the marker, the ancestral allele at that location in that generation, the frequency of that allele and the population in which it was recorded (1 or 2). If a critical fst value was used to terminate the simulation, and object FST with the final FST estimate is returned as well.

simulation\_data\_to\_genomeadmixr\_data
 function to convert ped/map data to genome\_admixr\_data

#### Description

function to convert ped/map data to genome\_admixr\_data

#### Usage

```
simulation_data_to_genomeadmixr_data(
   simulation_data,
   markers = NA,
   verbose = FALSE
)
```

#### Arguments

simulation\_data

	result of simulate_admixture
markers	vector of locations of markers (in Morgan). If no vector is provided, the function searches for marker locations in the simulation_data.
verbose	provide verbose output (default is FALSE)

#### Value

genomeadmixr\_data object ready for simulate\_admixture\_data

vcfR\_to\_genomeadmixr\_data

function to convert a vcfR object to genome\_admixr\_data

#### Description

function to convert a vcfR object to genome\_admixr\_data

#### Usage

```
vcfR_to_genomeadmixr_data(
  vcfr_object,
  chosen_chromosome,
  number_of_snps = NA,
  random_snps = TRUE,
  verbose = FALSE
)
```

#### Arguments

vcfr_object	result of vcfR::read.vcfR		
chosen_chromosome			
	chromosome of choice		
number_of_snps	number of snps to be loaded from the vcf file, default is to load all snps		
random_snps	if a subset of all snps has to be taken, should these be sampled sequentially (e., the first 100 snps) or randomly (100 randomly sampled snps) (examples are for 'number_of_snps' = 100).		
verbose	if true, print progress bar		

#### Value

genomeadmixr\_data object ready for simulate\_admixture\_data

write_as_plink	function to write simulation	n output as PLINK style data

#### Description

function to write simulation output as PLINK style data

#### Usage

```
write_as_plink(
    input_pop,
    marker_locations,
    file_name_prefix,
    chromosome = 1,
    recombination_rate = 1
)
```

#### Arguments

input\_pop input population, either of class "population" or of class "genomeadmixr\_data"
marker\_locations

location of markers, in bp

```
file_name_prefix
```

prefix of the ped/map files.

chromosome chromosome indication for map file

recombination\_rate

recombination rate in cM / kb

#### Value

No return value

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